

Total Syntheses of Natural Tubelactomicins B, D, and E: Establishment of Their Stereochemistries[†]

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Total syntheses of the antimicrobial tricyclic 16-membered macrolides, (+)-tubelactomicin B, (+)-tubelactomicin D, and (+)-tubelactomicin E, have been accomplished for the first time with common synthetic approaches. These total syntheses established the relative and absolute configurations of the three tubelactomicins, for which planar structures had solely been reported. The total synthesis of (+)-tubelactomicin D included a newly developed stereoselective intramolecular Diels—Alder reaction for constructing the highly functionalized octahydronaphthalene substructures.

Introduction

(+)-Tubelactomicin A (1) (Figure 1) was isolated from the culture broth of an actinomycete strain designated MK703-102F1, and its structure was determined by the researchers at the Institute of Microbial Chemistry in Japan. 1b This tricyclic 16-membered macrolide **1** showed potent antimicrobial activity against acid-fast bacteria, including drug-resistant strains. la Following the isolation of 1, the same group isolated and characterized structurally similar macrolides, i.e., (+)-tubelactomicin B (2), (+)-tubelactomicin D (3), and (+)-tubelactomicin E (4), from the same microoganism.² These antibiotics 2-4 also showed a broad range of antimicrobial activity. The planar structures of 2-4 were determined on the basis of spectral analysis [UV, IR, ¹H and ¹³C NMR]. Although their relative and absolute stereochemistries remained undetermined, it was considered that 2-4 possess the same stereochemistries as 1. In 2005, we reported the total synthesis of 1, thereby establishing the stereochemistry of the (+)-natural form.³ Tatsuta and coworkers have also accomplished the total synthesis of 1.4 Herein, we report the total syntheses of three other tubelactomicins, 2-4,

FIGURE 1. Structures of tubelactomicins.

thereby establishing their unknown relative and absolute stereochemistries as those depicted in Figure 1.

[†] This paper is dedicated to the memory of the late Professor Yoshihiko Ito. (1) (a) Igarashi, M.; Hayashi, C.; Homma, Y.; Hattori, S.; Kinoshita, N.; Hamada, M.; Takeuchi, T. *J. Antibiot.* **2000**, *53*, 1096–1101. (b) Igarashi, M.; Nakamura, H.; Takeuchi, T. *J. Antibiot.* **2000**, *53*, 1102–1107.

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SCHEME 1. Retrosynthetic Approach to Tubelactomicins B (2) and D (3)

We envisioned that the total syntheses of 2-4 would be accomplished using convergent synthetic approaches similar to that used for the total synthesis of 1. Therefore, the targeted macrolides 2 and 3 were divided into two segments, i.e., the upper-half segment 5 (for 2) or 6^{3b} (for 3) and the lower-half segment 7^{3a} (for 2) or 8 (for 3) as shown in Scheme 1. The novel upper-half segment 5 would be synthesized from 9, which was prepared previously from methyl (R)-lactate, 3b using the analogous carbon-carbon bond extension strategy including the Evans syn-aldol strategy⁵ to that used for the synthesis of another upper-half segment 6. The novel highly oxygenated octahydronaphthalene derivative 8, as the lower-half segment of 3, would be synthesized through an endo- and π -facial selective intramolecular Diels-Alder (IMDA) reaction⁶ of unsaturated aldehyde carrying a dienyne segment, i.e., 10. We previously observed that the structurally similar octahydronaphthalene derivative 7 was obtained stereoselectively by the IMDA

SCHEME 2. Synthesis of the Upper-Half Segment 5

reaction of the substrate with a methyl group instead of the (methoxymethoxy)methyl (MOM) group in 10.3a This substrate 10 for the attempted IMDA reaction would be obtained by the (E)-selective Horner—Wadsworth—Emmons olefination of highly oxygenated hexanal 11 and a novel (E)-trisubstituted olefin 12, which possesses a (trimethylsilyl)acetylene group, a diethyl methylphosphonate moiety, and an MOM-protected hydroxylmethyl group. The aldehyde 11 was previously prepared from diethyl (R)-malate^{3a} via a stereoselective α -allylation developed by Seebach and co-workers.⁷ As established in the case of the total synthesis of 1,3b the upper- and lower-half segments, i.e., the combination of 5 and 7 or 6 and 8, might be connected via a stereoselective sp²-sp² coupling, followed by a 16-membered macrolactonization, to construct the entire framework of 2 or 3, respectively. On the other hand, the known trans-fused octahydronaphthalene derivative, 3a which was synthesized as a key intermediate for the total synthesis of 1, could be used for the total synthesis of another macrolide 4 as the coupling partner of 6 (vide infra). Following these synthetic plans, we embarked on the total syntheses of 2-4.

Results and Discussion

The synthesis of **5** from the known **9** is summarized in Scheme 2. Compound **9** was converted to α -methylated α,β -unsaturated ester **14** by diisobutylaluminum hydride (DIBAL-H) reduction followed by a Wittig olefination of the

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SCHEME 3. Synthesis of the Allylic Phosphonate 12

resulting aldehyde 13. The ester 14 was converted to unsaturated aldehyde 16 by a reduction-oxidation protocol via allylic alcohol 15. The highly syn-selective boron-mediated Evans aldol reaction of 16 and an N-propionyl Evans chiral oxazolidinone⁵ produced the syn-adduct 17 with excellent stereoselection. 3b The hydroxyl group in the aldol product 17 was protected as the MOM ether, and the chiral auxiliary was reductively removed from the resulting 18 to produce the primary alcohol 19. Oxidation of 19 with Dess-Martin periodinane (DMP),8 followed by Corey-Fuchs dibromoolefin formation⁹ of the resulting aldehyde 20, produced α,α -dibromoethylene derivative 21 efficiently. Base-induced elimination of hydrogen bromide from 21 provided bromoalkyne 22. Deprotection of the tert-butyldiphenylsilyl (TBDPS) group in 22 with HF·pyridine, followed by treatment of the resulting 23 with tributylstannane in the presence of a catalytic amount of Pd(PPh₃)₄, ¹⁰ provided (E)vinylstannane 5 stereoselectively.

For the synthesis of the lower-half segment **8**, we first prepared allylic phosphonate **12** from the α -hydroxymethylated methyl acrylate **24**, ¹¹ the Morita—Baylis—Hillman product of methyl acrylate and p-formaldehyde. The synthesis of **12** from **24** is shown in Scheme 3. The allylic alcohol **24** was protected as the MOM ether. The addition of bromine to the MOM ether **25** produced dibromide **26** in 66% yield. The de-O-MOM derivative **27** was also produced, which was readily separated from **26**.

Treatment of **26** with tetrabutylammonium fluoride (TBAF) in HMPA provided (*E*)-bromoalkene **28** stereoselectively as a result of hydrogen bromide elimination from **26**. The Pd (0)-catalyzed Sonogashira coupling of **28** and (trimethylsilyl)-acetylene in the presence of CuI and *i*-Pr₂NEt¹² produced enyne ester **29** in high yield. The DIBAL-H reduction of **29** and allylic

SCHEME 4. Synthesis of the Substrate 10 and the IMDA Reaction

bromination of the resulting allylic alcohol 30 with carbon tetrabromide in the presence of triphenylphosphine provided allylic bromide 31. The Arbusov rearrangement of 31 with triethyl phosphite produced functionalized diethyl allylphosphonate 12.

The synthesis of 8, commencing with the Horner-Wadsworth-Emmons reaction of the known aldehyde 113a and allylphosphonate 12, is shown in Schemes 4 and 5. From the coupling of 11 and 12, the (E,E)-dienyne-type adduct 32 was obtained stereoselectively. Acid hydrolysis of the silyl ether in 32, oxidation of the resulting 33 with DMP, followed by the olefination of aldehyde 34 with Ph₃P=C(Me)CO₂Et produced α,β -unsaturated ester 35 with exclusive (E)-selectivity. The (E)geometry of the unsaturated ester moiety of 35 was secured by NOE experiment in the ¹H NMR analysis of 35. Thus a 10% signal enhancement of the α -methyl group in the α,β -unsaturated ester in 35 was observed when one of the allylic methylene protons was irradiated. A reduction-oxidation strategy was used for the conversion of 35 to the IMDA substrate 10. Thus, the DIBAL-H reduction of 35 provided allylic alcohol 36, which was oxidized with DMP to produce 10. The attempted IMDA reaction of 10 proceeded at 80 °C in toluene, as observed in the IMDA reaction of the less congested substrate, which possessed a methyl group instead of the MOMOCH2 group in the diene part of 10.3a The IMDA reaction produced two adducts 37 and 38 as an inseparable mixture. Thus, the mixture was further oxidized to the respective carboxylic acids 39 and 40 by a perchlorite oxidation. The carboxylic acids 39 and 40 were cleanly separated by chromatography on silica gel in 76% and 18% yield, respectively, for the two steps. The stereochemistry

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2 (83%)

SCHEME 5. Conversion of 39 to the Lower-Half Segment 8

of the major product **39** was determined to be the desired *endo*-adduct unambiguously by 1 H NMR analysis including detailed NOE experiments. For the stereochemistry of the minor product **40**, we assigned the *exo*-stereochemistry as depicted in Scheme 4 on the basis of our previous result experienced for the total synthesis of **1**. 3a In the previous case, 3a the ratio of the *endo*-and *exo*-adduct was approximately 8 to 1. In the present case, the *endo/exo* selectivity of the IMDA reaction of **10** was estimated to be approximately 4:1. It is apparent that the bulkiness of the more congested group in the diene part diminishes the *endo/exo* ratio of the IMDA reaction. As we observed in the case of **1**, the π -facial selectivity in the IMDA reaction of **10** was completely controlled.

Having achieved the synthesis of the desired IMDA adduct 39, we explored the conversion of 39 to the lower-half segment 8 as shown in Scheme 5. Protodesilylation of the *C*-TMS group in 39 with TBAF and hydrolytic de-*O*-benzylidenation, followed by (2-trimethylsilylethoxy)methyl (SEM) ester formation, provided 43 via 41 and 42. Selective tosylation of diol 43 and deoxygenation of the resulting primary tosylate 44 with NaBH₄ in hot DMSO provided 45. Regio- and *cis*-selective palladium-catalyzed hydrostannylation of the acetylene functionality in 45 and subsequent metal—iodine exchange of the resulting vinyl-stannane provided the lower-half segment 8 in good overall yield.

We then explored the Stille coupling¹³ of **5** and **7** and also that of **6** and **8** for the total syntheses of **2** and **3**, respectively. The reaction conditions and results of the Stille couplings and transformation of the coupling products **46** and **49** into **2** and **3**, respectively, are summarized in Scheme 6. The palladium-catalyzed Stille coupling of **5** and **7** proceeded with CuI in the presence of AsPh₃ in DMF, ¹⁴ providing (*E,E*)-diene **46** efficiently. Deprotection of the SEM ester in **46** with HF•pyridine and lactonization of the resulting seco acid **47** under the Mukaiyama conditions (2-chloro-1-methylpyridinium iodide and triethylamine) ¹⁵ provided 16-membered macrolactone **48** in an

SCHEME 6. Completion of the Total Syntheses of 2 and 3

52

6 M aqueous HCI/THF (1:1)

48

excellent yield of 93%. Finally, acid hydrolysis of the MOM group in 48 provided (+)-tubelactomicin B (2), which was found to be identical with a natural sample by comparison of spectra (1 H and 13 C NMR, IR), [α]_D, and TLC behavior. The dextrorotatory property of synthetic 2 proved that the absolute stereochemistry of natural 2 was as depicted. Similarly, the Stille coupling of 6 and 8 provided (*E*,*E*)-diene 49 under the same conditions as those used for the coupling of 5 and 7. The Mukaiyama macrolactonization of the seco acid 50 derived from 49 provided 51 with a lower yield of 59% for two steps. The two-step deprotection of 51 via 52 eventually provided (+)-tubelactomicin D (3), which was identical with a natural sample ([α]_D, 1 H and 13 C NMR, IR, and TLC behavior), establishing the absolute configuration of 3.

The total synthesis of (+)-tubelactomicin E (4) was started with the previously reported intermediate 53^{3a} used for the total synthesis of 1. The total synthesis of 4 from 53 is shown in Scheme 7. The octahydronaphthalene derivative 53 was in turn synthesized featuring an *endo*- and π -facial selective IMDA reaction of a D-malic acid-derived substrate. According to the conversion of 45 to 8, the carboxylic acid 53 was esterified with SEMCl, and the resulting SEM ester 54 was subjected to the palladium-catalyzed hydrostannylation, followed by tin—iodine exchange, providing 55. The palladium-catalyzed Stille coupling of the upper-half segment 6^{3b} and vinyl iodide 55 provided conjugated (E,E)-diene 56. Deprotection of the SEM group in 56 and the Mukaiyama macrolactonization of the resulting seco acid 57 provided 58, from which (+)-tubelacto-

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SCHEME 7. Total Synthesis of (+)-Tubelactomicin E (4)

micin E (4) was synthesized by a two-step deprotection via **59**. The synthetic **4** was identical with a natural sample ($[\alpha]_D$, 1H and ^{13}C NMR, IR, and TLC behavior).

In conclusion, we have completed the total syntheses of natural (+)-tubelactomicins B (2), D (3), and E (4) for the first time. These total syntheses also established the relative and absolute stereochemistries of these natural products, which had not been determined.

Experimental Section

(1E,3S,4R,5E,10R)-10-Hydroxy-4-methoxymethoxy-3,5-dimethyl-1-(tributylstannyl)undeca-1,5-diene (5). The following reaction was carried out under argon. To a stirred solution of 23 (353 mg, 1.06 mmol) in THF (7 mL) were added Pd(PPh₃)₄ (123 mg, 0.10 mmol) and Bu₃SnH (0.85 mL, 3.16 mmol). The reaction mixture was stirred for 10 min and then concentrated in vacuo. The residue was purified by column chromatography on silica gel (EtOAc/hexane, 1:4 doped with Et₃N) to give 504 mg (87%) of 5 as a colorless oil. ¹H NMR analysis revealed the product contained approximately 9% of the Z-isomer. This mixture was used for subsequent steps without separation. Compound 5: TLC, R_f 0.48 (EtOAc/hexane, 1:2); $[\alpha]^{22.5}_{D} + 58.7$ (c 0.85, CHCl₃); ¹H NMR (300) MHz) δ 0.78–0.95 (m, 15H), 1.11 (d, J = 6.6 Hz, 3H), 1.18 (d, J= 6.6 Hz, 3H, 1.22 - 1.38, 1.40 - 1.54 (m, 12H), 1.22 - 1.54 (m, 12H)4H), 1.48 (s, 3H), 1.60 (s, 1H), 1.88 (m, 2H), 2.41 (ddq, J = 7.1, 9.0, 6.2 Hz, 1H), 3.38 (s, 3H), 3.63 (d, J = 9.0 Hz, 1H), 3.84 (m, 1H), 4.45 (d, J = 6.6 Hz, 1H), 4.62 (d, J = 6.6 Hz, 1H), 5.32 (t, J = 6.6 Hz, 1H), 5.71 (dd, J = 7.1, 19.0 Hz, 1H), 5.87 (d, J = 19.0 Hz, 1H); ^{13}C NMR (68 MHz) δ 9.4 \times 3, 11.4, 13.7 \times 3, 17.1, 23.4, 25.7, 27.2, 27.4 \times 3, 29.1 \times 3, 39.1, 44.2, 55.6, 67.9, 85.8, 93.0, 126.5, 130.3, 132.9, 150.9; IR (neat) 3400, 2960, 1600, 1460 cm $^{-1}$; HRMS calcd for $C_{23}H_{45}O_{3}Sn$ (M $^{+}-$ Bu) $\emph{m/z}$ 489.2391, found 489.2392.

2-(Trimethylsilyl)ethoxymethyl (1R,2S,4aR,5R,6S,8aS)-5-Hydroxy-2-[(1E,3E,5S,6R,7E,12R)-12-hydroxy-6-methoxymethoxy-5,7-dimethyltrideca-1,3,7-trienyl]-1,3,6-trimethyl-1,2,4a,5,6,7,8,-8a-octahydronaphthalene-1-carboxylate (46). The following reaction was carried out under argon. To a stirred solution of 5 (55.4 mg, 102 mmol) and 7^{3a} (38.5 mg, 74.0 mmol) in degassed DMF (1 mL) was added a solution of Pd₂(dba)₃ (3.5 mg, 3.8 mmol), AsPh₃ (9.3 mg, 30 mmol), and CuI (2.9 mg, 15 mmol) in degassed DMF (0.25 mL). The mixture was stirred for 45 min and then heated at 60 °C for 2 h. After being cooled to room temperature, the mixture was diluted with saturated aqueous NaHCO₃ (20 mL) and extracted with Et₂O (3 \times 10 mL). The combined organic layers were dried and concentrated with the aid of toluene. The residue was purified by column chromatography on silica gel (EtOAc/ hexane, 1:5 to 1:3) to give 35.8 mg (75%) of **46** as a colorless oil: TLC, R_f 0.66 (EtOAc/hexane, 2:1); $[\alpha]^{26.5}$ _D +216 (c 0.55, CHCl₃); ¹H NMR (300 MHz) δ 0.03 (s, 9H), 0.96 (t, J = 8.3 Hz, 2H), 1.05 (d, J = 6.4 Hz, 3H), 1.10 (d, J = 6.6 Hz, 3H), 1.16 (s, 3H), 1.18(d, J = 6.2 Hz, 3H), 1.32-1.51 (m, 5H), 1.49 (s, 3H), 1.62 (s,3H), 1.60-1.83 (m, 6H), 1.96-2.09 (m, 2H), 2.35 (d, J = 9.6 Hz, 1H), 2.30-2.47 (m, 1H), 2.91 (m, 1H), 3.37 (s, 3H), 3.57 (d, J =9.0 Hz, 1H), 3.68 (t, J = 8.3 Hz, 2H), 3.78 (m, 1H), 4.44, 4.60 $(2d, J = 6.6 \text{ Hz}, 1H \times 2), 5.10, 5.21 (2d, J = 6.0 \text{ Hz}, 1H \times 2),$ 5.28 (m, 1H), 5.29 (t, J = 6.8 Hz, 1H), 5.44 (m, 1H), 5.76 (s, 1H), 5.82-5.98 (m, 2H); 13 C NMR (68 MHz) δ -1.4 × 3, 11.3, 16.5, 16.8, 18.0, 18.7, 22.6, 23.5, 25.6, 27.0, 27.4, 33.3, 37.7, 39.0, 40.8. 45.5, 49.7, 54.3, 55.6, 67.8, 67.9, 79.5, 85.8, 89.5, 93.0, 121.0, 128.9, 130.6, 131.3, 132.4, 132.6, 133.0, 135.6, 175.1; IR (neat) 3420, 2950, 2930, 2880, 1730, 1710, 1460 cm⁻¹; HRMS calcd for $C_{37}H_{64}O_7Si$ (M⁺) m/z 648.4421, found 648.4419.

(1R,2S,4aR,5R,6S,8aS)-5-Hydroxy-2-[(1E,3E,5S,6R,7E,12R)-12-hydroxy-6-methoxymethoxy-5,7-dimethyltrideca-1,3,7-trienyl]-1,3,6-trimethyl-1,2,4a,5,6,7,8,8a-octahydronaphthalene-1carboxylic Acid (47). To a cooled (0 °C), stirred solution of 46 (35.5 mg, 54.7 mmol) in THF (1 mL) was added HF+pyridine complex (0.2 mL). The mixture was stirred for 21 h and quenched with saturated aqueous NaHCO₃ (4 mL) at 0 °C. This was diluted with saturated aqueous NaHCO₃ (16 mL) and extracted with CH_2Cl_2 (3 × 20 mL). The combined organic layers were dried and concentrated with the aid of toluene. The residue was purified by column chromatography on silica gel (EtOAc/hexane, 1:3) to give 24.3 mg (86%) of **47** as a colorless oil: TLC, R_f 0.51 (EtOAc/ hexane, 2:1); $[\alpha]^{25}_D$ +198 (c 1.02, CHCl₃); ¹H NMR (300 MHz) δ 1.04 (d, J = 6.4 Hz, 3H), 1.14 (d, J = 6.8 Hz, 3H), 1.15 (s, 3H), 1.19 (d, J = 6.2 Hz, 3H), 1.31 - 1.49 (m, 5H), 1.51 (s, 3H), 1.63(s, 3H), 1.55-1.83 (m, 6H), 1.93-2.13 (m, 2H), 2.38 (d, J=9.4Hz, 1H), 2.32-2.49 (m, 1H), 2.91 (m, 1H), 3.37 (s, 3H), 3.57 (d, J = 9.4 Hz, 1H), 3.77–3.92 (m, 1H), 4.44, 4.60 (2d, J = 6.7 Hz, $1H \times 2$), 5.28-5.52 (m, 3H), 5.75 (s, 1H), 5.85-6.02 (m, 2H); 13 C NMR (68 MHz) δ 11.3, 16.6, 16.7, 18.7, 22.8, 23.1, 25.3, 26.8, 27.1, 33.5, 37.6, 38.3, 38.4, 40.8, 45.7, 49.0, 53.9, 55.6, 68.3, 79.5, 86.2, 92.9, 120.9, 129.3, 130.8, 131.5, 132.4, 132.6, 133.2, 134.7, 178.3; IR (neat) 3420, 2970, 2930, 2880, 1710, 1690, 1460, 1450 cm^{-1} .

(15,2E,4E,6S,7R,8E,13R,16R,17S,20S,21R,22R)-21-Hydroxy-7-methoxymethoxy-6,8,13,16,20,24-hexamethyl-14-oxatricyclo-[14.8.0.0^{17,22}]tetracosa-2,4,8,23-tetraene-15-one (48). A solution of 47 (20.2 mg, 38.9 μ mol) in MeCN (24 mL) was added to a refluxing solution of 2-chloro-1-methylpyridinium iodide (50.1 mg, 0.196 mmol) and Et₃N (0.055 mL, 0.40 mmol) in MeCN (12 mL) over 2.5 h by an addition funnel. The addition funnel was rinsed with 3.5 mL of MeCN and the washing was added to the reaction mixture. The mixture was refluxed for an additional 3.5 h. After being cooled to room temperature, the mixture was concentrated



in vacuo. The residue was purified by column chromatography on silica gel (EtOAc/hexane, 1:4) to give 18.2 mg (93%) of 48 as a colorless oil: TLC, R_f 0.76 (EtOAc/hexane, 1:1); $[\alpha]^{26}_D$ +182 (c 0.875, CHCl₃); ¹H NMR (300 MHz) δ 0.83-0.98 (m, 2H), 1.04 (d, J = 6.2 Hz, 3H), 1.12 (s, 3H), 1.16-1.29 (m, 6H), 1.34-1.50(m, 2H), 1.52 (s, 3H), 1.57-1.60 (m, 7H), 1.63 (s, 3H), 1.96-2.24 (m, 2H), 2.34 (d, J = 10.2 Hz, 1H), 2.39 - 2.51 (m, 1H), 2.92(m, 1H), 3.37 (s, 3H), 3.48 (d, J = 9.6 Hz, 1H), 4.44, 4.58 (2d, J $= 6.4 \text{ Hz}, 1\text{H} \times 2), 4.62 - 4.76 \text{ (m, 1H)}, 5.09 \text{ (m, 1H)}, 5.31 \text{ (dd, } J$ = 14.1, 10.2 Hz, 1H), 5.47 (dd, J = 14.9, 6.5 Hz, 1H), 5.73 (s, 1H), 5.81–6.10 (m, 2H); 13 C NMR (68 MHz) δ 11.2, 15.7, 16.7, 18.7, 19.2, 22.6, 25.3, 26.4, 26.8, 33.3, 34.8, 36.7, 38.1, 40.9, 45.6, 48.5, 54.3, 55.5, 71.2, 79.6, 88.1, 93.0, 120.5, 127.8, 130.9, 131.8, 132.8, 133.4 × 2, 135.5, 174.2; IR (neat) 3490, 2980, 2930, 2880, 1720, 1460 cm⁻¹; HRMS (FAB) calcd for $C_{31}H_{48}O_5$ (M⁺) m/z500.3502, found 500.3505.

(+)-Tubelactomicin B (2). To a cooled (0 °C), stirred solution of 48 (17.5 mg, 34.9 µmol) in THF (0.5 mL) was added 6 M aqueous HCl (0.5 mL). The mixture was stirred for 11 h and then additional 6 M aqueous HCl (0.25 mL) and THF (0.5 mL) were added. The mixture was stirred for an additional 6 h and neutralized with saturated aqueous NaHCO3 (15 mL) at 0 °C. This was extracted with CH₂Cl₂ (3 × 8 mL). The combined organic layers were dried and concentrated in vacuo. The residue was purified by column chromatography on silica gel (EtOAc/hexane, 1:4) to give 13.3 mg (83%) of **2** as a colorless solid: TLC, R_f 0.58 (CHCl₃/ MeOH, 10:1), R_f 0.62 (EtOAc/hexane, 1:1); $[\alpha]^{24.5}_D$ +101(c 0.60, MeOH); ¹H NMR (300 MHz) δ 0.83–1.00 (m, 2H), 1.04 (d, J =6.2 Hz, 3H), 1.12 (s, 3H), 1.18 (d, J = 6.8 Hz, 3H), 1.21 (d, J =6.2 Hz, 3H), 1.32-1.51 (m, 2H), 1.60 (s, 3H), 1.61-1.88 (m, 7H), 1.63 (s, 3H), 1.93-2.20 (m, 2H), 2.32-2.45 (m, 1H), 2.35 (d, J =10.2 Hz, 1H), 2.92 (m, 1H), 3.57 (d, J = 9.0 Hz, 1H), 4.68 (m, 1H), 5.06 (m, 1H), 5.31 (dd, J = 14.0, 10.2 Hz, 1H), 5.48 (dd, J= 15.0, 6.4 Hz, 1H), 5.73 (s, 1H), 5.81–5.99 (m, 2H); 13 C NMR (68 MHz) δ 11.1, 15.5, 16.7, 18.7, 19.2, 22.6, 25.3, 26.3, 26.8, 33.3, 34.8, 38.1, 40.9, 45.6, 48.5, 54.3, 71.3, 79.6, 85.3, 120.5, 127.6, 129.2, 130.8, 133.3, 133.4, 135.5, 136.2, 174.3; IR (KBr) 3420, 2980, 2930, 2880, 1720, 1705, 1460 cm⁻¹; HRMS calcd for C₂₉H₄₄O₄ (M⁺) m/z 456.3240, found 426.3234.

(+)-**Tubelactomicin D** (3). To a stirred solution of **52** (40.1 mg, 77.6 μmol) in MeOH (3 mL) was added 1 M aqueous NaOH (1.5 mL). The mixture was heated at 50 °C with stirring for 6.5 h. After being cooled to 0 °C, the mixture was acidified with 1 M aqueous HCl to pH 2. This was diluted with H₂O (4 mL) and extracted with CH₂Cl₂ (3 × 3 mL). The combined organic layers were dried and concentrated in vacuo. The residue was purified by column chromatography on silica gel (hexane/acetone, 2:1) to give **3** (36.0 mg, 92%) as a colorless oil: TLC, R_f 0.22 (CHCl₃/MeOH, 10:1 containing 1% AcOH); [α]^{24.5}_D +95.7 (c 0.14, CHCl₃); ¹H NMR (300 MHz, acetone- d_6) δ 1.03 (d, J = 6.4 Hz, 3H), 1.12 (s, 3H), 1.15 (d, J = 6.0 Hz, 3H), 1.21 (d, J = 7.3 Hz, 3H), 1.27–1.33 (m, 3H), 1.34–1.71 (m, 3H), 1.60–1.78 (m, 4H), 1.84–1.88 (m, 1H),

2.40–2.60 (m, 2H), 2.72 (d, J=10 Hz, 2H), 2.84 (t, J=9.0 Hz, 2H), 3.89–3.93 (m, 5H), 4.71 (m, 1H), 5.28 (dd, J=10, 14 Hz, 1H), 5.72 (dd, J=5.7, 14 Hz, 1H), 5.81 (dd, J=4.6, 8.3 Hz, 1H), 5.89–6.00 (m, 2H), 6.09 (br s, 1H); ¹³C NMR (75 Hz, acetone- d_6) δ 15.9, 17.1, 19.4, 19.5, 26.1, 27.8, 29.0, 34.4, 35.9, 39.6, 41.2, 41.9, 46.5, 48.9, 50.5, 65.1, 71.4, 79.4, 81.8, 122.9, 128.7, 131.7, 134.3, 134.7, 137.0, 137.7, 144.7, 168.6, 174.4; IR (neat) 3600, 3500, 2980, 1740, 1680, 1650, 1460 cm⁻¹; HRMS calcd for $C_{29}H_{42}O_7$ (M⁺) m/z 502.2930, found 502.2929.

(+)-Tubelactomicin E (4). To a stirred solution of 59 (9.9 mg, 19 µmol) in MeOH (1 mL) was added 1 M aqueous NaOH (0.5 mL). The mixture was heated to 50 °C for 3 h. After being cooled to room temperature, the mixture was acidified with 1 M aqueous HCl to pH 2 at 0 °C. This was diluted with 1 M aqueous HCl (10 mL) and extracted with CH_2Cl_2 (3 × 5 mL). The combined organic layers were dried and concentrated in vacuo. The residue was purified by column chromatography on silica gel (acetone/hexane, 1:1 containing 1% AcOH) to give 9.1 mg (94%) of 4 as a colorless solid: TLC, R_f 0.27 (CHCl₃/MeOH, 10:1 containing 1% AcOH), 0.36 (acetone/hexane, 1:1 containing 1% AcOH); $[\alpha]^{23.5}_{D}$ +94.9 (c 0.23, MeOH); ¹H NMR (300 MHz, acetone- d_6) δ 1.01–1.76 (m, 10H), 1.13 (s, 3H), 1.15 (d, J = 6.0 Hz, 3H), 1.23 (d, J = 6.3Hz, 3H), 1.61 (s, 3H), 1.84 (m, 1H), 2.40 (d, J = 10.0 Hz, 1H), 2.47-2.57 (m, 2H), 2.60-2.73 (m, 1H), 3.22 (dd, J = 9.3, 9.3 Hz, 1H), 3.65 (dd, J = 10.5, 6.6 Hz, 1H), 3.72 (dd, J = 10.5, 5.1 Hz, 1H), 3.94 (d, J = 8.4 Hz, 1H), 4.71 (m, 1H), 5.29 (dd, J = 13.9, 10.0 Hz, 1H), 5.70 (dd, J = 14.6, 6.2 Hz, 1H), 5.83 (s, 1H), 5.83 6.00 (m, 3H); 13 C NMR (75 MHz, acetone- d_6) δ 16.1, 17.2, 19.4, 22.9, 26.0, 27.2, 28.1, 35.8, 38.5, 41.9, 46.5, 48.1, 49.1, 55.1, 67.1, 71.4, 77.4, 81.3, 122.3, 128.9, 131.8, 133.0, 134.2, 134.9, 136.9, 144.4, 168.6, 174.4 (solvent peak overlapped one carbon peak, so one carbon was not detected); IR (KBr) 3420, 2930, 2860, 1720, 1700, 1460 cm⁻¹; HRMS calcd for $C_{29}H_{42}O_7$ (M⁺) m/z 502.2930, found 502.2930.

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Supporting Information Available: Experimental procedures and full spectroscopic data for all new compounds described herein and copies of ¹H and ¹³C NMR spectra for all new compounds, synthetic and natural tubelactomicins B, D, and E. This material is available free of charge via the Internet at http://pubs.acs.org.

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